

Impact of baseline molecular alterations on the efficacy of tucatinib (TUC) plus trastuzumab (Tras) for HER2+, RAS WT metastatic CRC (mCRC) in MOUNTAINEER

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DECLARATION OF INTERESTS

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Background

- HER2 overexpression/amplification occurs in 3-5% of patients presenting with mCRC¹⁻⁵
- Established regional guidelines for mCRC recommend HER2 testing and HER2-directed treatment options^{1,6}
- Tucatinib is a highly selective HER2-directed TKI approved by the FDA in combination with trastuzumab for treatment of patients with RAS wild-type HER2+ unresectable or mCRC that has progressed following treatment with fluoropyrimidine-, oxaliplatin-, and irinotecanbased chemotherapy⁷
- Treatment response to TUC+Tras was predicted by a HER2+ result from IHC, ISH, or NGS testing⁸
- Concomitant genomic alterations are common in patients with HER2+ mCRC⁹ but the impact of these alterations on clinical outcomes is unknown

 Benson AB, J Natl Compr Canc Netw. 2021: 329-59. 2. Kang A. J Manag Care Spec Pharm. 2021: S20-1. 3. Patel JN. J Precis Med. 2019: 3. 4. Sartore-Bianchi A. Oncologist. 2019: 1395-402. 5. Strickler J. J Clin Oncol. 2021: Abstract TPS153. 6. Cervantes A. Ann Oncol. 2023: 10-32. 7. TUKYSA. Prescribing Information. Seagen Inc. Jan 2023. Accessed Sep 11, 2023. 8. Ucital Viciente J. 2019. 10(1): Oncol. 2023;41(Suppl 16):Abstract 3528. 9. Cancer Genome Atlas Network. Nature. 2012;487(7407): 330-IHC, immunohistochemistry; ISH, in situ hybridization; mCRC, metastatic colorectal cancer; NGS, next-generation sequencing; TKI, tyrosine kinase inhibitor



MOUNTAINEER

Results: Cohorts A+B¹

38 1%

12.4 months



^a Tucatinib dose: 300 mg PO BID, trastuzumab dose: 6 mg/kg Q3W (loading dose 8 mg/kg C1D1); each treatment cycle is 21 days

^b Patients remained on therapy until evidence of radiographic or clinical progression, unacceptable toxicity, withdrawal of consent, or study closure

^c Stratification: left sided tumor primary vs other

^d Patients were allowed to cross over and receive tucatinib and trastuzumab if they experienced radiographic progression at any time point or if they had not achieved a PR or CR by Week 12

e Efficacy assessed in patients who received any amount of study treatment and had HER2+ tumors

mPFS 8.2 months mOS 24.1 months Safety (n=86)

Diarrhea was most common AE

- Grade 1 or 2: 60.5%
- Grade 3: 3.5%

Efficacy (n=84)

cORR per BICR

mDOR

Results from the MOUNTAINEER primary analysis showed clinically meaningful activity and demonstrated tucatinib in combination with trastuzumab was well tolerated¹

¹Strickler JH. Lancet Oncol. 2023: 496-508.

AE, adverse event; BICR, blinded independent central review; BID, twice a day; C1D1, Cycle 1 Day 1; cORR, confirmed objective response rate; CR, complete response; IHC, immunohistochemistry; ISH, in situ hybridization; DOR, duration of response; m, median; mAb, monoclonal antibody; NGS, next-generation sequencing;

OS, overall survival; PFS, progression-free survival; PO, orally; PR, partial response; Q3W, every 3 weeks; RECIST, Response Evaluation Criteria in Solid Tumors

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f RECIST v1.1 per BICR

Exploratory Biomarker Analyses



^aConfirmed HER2-positive mCRC tested at CLIA-certified or ISO-accredited laboratory: IHC 3+ or 2+ with (F)ISH reflex by an FDA-approved or CE-marked test

^bAnalysis performed by sponsor-designated central laboratory

ePatients were required to have confirmed HER2 positive status via a retrospective HER2+ central lab test from one of the following methods: IHC/FISH, tissue NGS, or ctDNA

^dcORR and DOR were calculated for SNVs and CNVs occurring in ≥5 patients; cORR was defined as the proportion of patients with complete response or partial response per RECIST v1.1 by BICR

BICR, blinded independent central review; CEN17, centromere of chromosome 17; CNV; copy number variant; cORR, confirmed objective response rate; ctDNA, circulating tumor DNA; DOR, duration of response; EOT, end of treatment; FISH, fluorescence in situ hybridization; IHC, immunohistochemistry; ISH, in situ hybridization; mCRC, metastatic colorectal cancer; NGS, next-generation sequencing; RECIST; Response Evaluation Criteria in Solid Tumors; SNV, single nucleotide variant



Results: Samples Available for Testing at Baseline



FISH testing utilized the gastric criteria for purposes of this analysis Final sample sizes for FISH, Tissue NGS, and ctDNA consist of patients with ≥1 positive central test result regardless of modality ctDNA, circulating tumor DNA; FISH, fluorescence in situ hybridization



cORR by Clinicopathologic features and Testing Method

	Full analysis set		Central FISH cORR		Central Tissue N	IGS cORR	Central ctDNA cORR	
		cORR, n/N %		cORR, n/N %		cORR, n/N %		cORR, n/N %
Overall	⊢ ∎-1	32/84 38.1	⊢ – −1	26/65 40.0	⊢	21/45 46.7	⊢ ∎1	26/66 39.4
Primary tumor site	e							
Left sided	⊢■⊣	30/71 42.3		24/53 45.3	⊢_ ∎1	20/38 52.6	⊢ = -1	24/54 44.4
All other primary sites ^a	├─■──┤	2/13 15.4	⊢ − − 1	2/12 16.7	⊢ − − − 1	1/7 14.3	├─■──┤	2/12 16.7
Prior anti-EGFR th	herapy							
Yes	⊢ ∎1	16/44 36.4	⊢ ∎	13/34 38.2	⊢	11/25 44.0	⊢ – – – – – – – – – – – – – – – – – – –	13/33 39.4
No	⊢	16/40 40.0	⊢=1	13/31 41.9	⊢	10/20 50.0	⊢ • 1	13/33 39.4
Prior anti-VEGF therapy								
Yes	F-■1	29/72 40.3	⊢ ■ - 1	23/54 42.6	⊢_ ∎1	18/36 50.0	⊢	24/56 42.9
No	⊢ – – – 1	3/12 25.0		3/11 27.3	├── ■	3/9 33.3	├─── ─┤	2/10 20.0
Prior lines of therapy in metasta or recurrent settin	atic g							
1 line	├ ── ─ ──┤	9/19 47.4	⊢	7/16 43.8	├──●	4/11 36.4		7/16 43.8
2 lines	⊢ ∎−−1	15/32 46.9	⊢ =	13/27 48.1	-	- 13/21 61.9	⊢ ■	12/26 46.2
≥3 lines	┝──■──┤	8/33 24.2	⊢	6/22 27.3	├── ■──┤	4/13 30.8	┝──■──┤	7/24 29.2
	0 20 40 60 80 cORR (%)	100	0 20 40 60 80 cORR (%)	100	0 20 40 60 80 cORR (%)	100	0 20 40 60 80 cORR (%)	100
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Tissue FISH Assay

Impact of baseline HER2/CEN17 ratio and HER2 copy number



DOR subgroups not shown had <5 responders and analyses were not performed

HER2 copy number of 9.45 used in Sartore-Bianchi A. Lancet Oncol. 2016;17(6):738-46; HER2/CEN17 ratio used in Raghav KPS. J Clin Oncol 41, 2023 (suppl4; abstr 140)

Error bars represent 95% Cls

CEN17, centromere of chromosome 17; cORR, confirmed objective response rate; DOR, duration of response; FISH, fluorescence in situ hybridization

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Tissue NGS: Impact of Baseline Co-Amplifications and Co-Mutations



cORR was calculated for genomic alterations with a minimum sample size of 5 subjects. DOR was not calculated for genomic alterations with <5 responders cORR, confirmed objective response rate; DOR, duration of response; NE, not evaluable; NGS, next-generation sequencing

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ctDNA NGS: Impact of Baseline Co-Amplifications and Co-Mutations

								DOR,
Category	Total	Responders	i				cORR (95% CI)	months (95% CI)
ctDNA NGS	66	26			_		39.4 (27.6-52.2)	12.4 (8.3-20.5)
Amplifications								
BRAF	7	1		1			14.3 (0.4-57.9)	-
CCND1	7	1					14.3 (0.4-57.9)	-
CDK6	10	3					30.0 (6.7-65.2)	-
EGFR	19	7					36.8 (16.3-61.6)	12.5 (4.2-NE)
ERBB2	57	23					40.4 (27.6-54.2)	12.4 (6.2-38.3)
PIK3CA	5	2					40.0 (5.3-85.3)	_
Selected Mutations*								
APC	42	18					42.9 (27.7-59.0)	15.3 (6.2-38.3)
ERBB2	15	7	_				47.1 (23.0-72.2)	11.4 (6.1-NE)
KRAS/NRAS	5	2		-			40.0 (5.3-85.3)	-
PIK3CA	7	2		-		-	28.6 (3.7-71.0)	-
				1	1	Ì		
			0 20	40	60	80	100	
cORR (%)								

cORR was calculated for genomic alterations with a minimum sample size of 5 subjects. DOR was not calculated for genomic alterations with < 5 responders

*Additional genes with >5 patients with mutations (gene, responders/total): CDK12 6/11; BRCA2 4/6; BRCA1 1/5; FBXW7 4/6; ATM 1/6. For CDK12, DOR (95% CI) is 12.5 (4.2-NE).2 patients had KRAS

mutations and 0/2 were responders; 3 patients had NRAS mutations and 2/3 were responders.

ctDNA, circulating tumor DNA; cORR, confirmed objective response rate; DOR, duration of response; NE, not evaluable; NGS, next-generation sequencing

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ctDNA NGS: Genomic Landscape of Acquired Alterations at Progression Timepoint or EOT

PR	16.5
PR	15.2
PR	13.1
PR	12.6
PR	12.5
PR	10.3
PR	8.3
PR	8.3
SD	8.1
PR	8.1
PR	8
PR	6.8
SD	4.4
SD	3.1
SD	2.8
SD	2.7
SD	2.7
SD	2.6
SD	2.6
SD	2.6
SD	2.1
PD	2.1
PD	1.7
PD	1.5
PD	1.1
Response	PFS

**KRAS G13C, G12C, I24N



n=31; 1 patient removed from analysis due to no detected alterations at baseline, leading to analysis set of 30; 23/30 showed alteration gains; 2/30 showed ERBB2 loss; 5/30 showed no alteration gains and no ERBB2 loss.



Note: a single BLUE or YELLOW box can represent multiple SNV/INDEL detections in the same gene

DDR: DNA Damage Response; EOT, end of treatment; PFS, progression-free survival; RTK: Receptor Tyrosine Kinase; SNV, single nucleotide variation



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Conclusions

- In this exploratory analysis, TUC+Tras had broad clinical activity in all clinicopathologic and genomic subgroups
- Earlier lines of treatment (≤2L), left sided primary tumors, higher gene copy number, and increased HER2/CEN17 ratio might be associated with greater clinical benefit additional study is needed
- In the baseline tissue and ctDNA NGS analysis, *RAS* and *ERBB2* mutations did not predict primary treatment resistance
- At the time of progression (EOT), ctDNA demonstrated the following patterns:
 - Most patients acquired multiple heterogeneous resistance alterations
 - Common acquired genomic alterations included *PIK3CA*, *RTKs*^a, and *KRAS/NRAS*
 - Loss of ERBB2 amplification was rare
- Understanding the genomic drivers of primary and acquired resistance may inform future therapeutic opportunities



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